Filling the Gaps in Drug Therapy

Hutchinson-Gilford progeria syndrome

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Abstract

The rare genetic autosomal dominant condition Hutchinson-Gilford progeria syndrome (HGPS) is characterized by a dramatic, rapid appearance of aging beginning in childhood. HGPS progression generates vascular disease, which generally leads to death during the teenage years. There is currently no treatment for this genetic disorder, although since the identification of the underlying genetic mutation, new targets and potential strategies have emerged. The large majority of HGPS cases carry a single nucleotide substitution within exon 11 of the LMNA gene and abnormal splicing results in production of a truncated prelamin A protein (progerin). Subsequent farnesylation of mutated prelamin A targets the protein to the nuclear envelope, resulting in nuclear deformations and blebbing. Recent studies have shown that blocking farnesylation of progerin via the use of farnesyltransferase inhibitors can reduce nuclear blebbing and thus HGPS pathogenicity. Lentiviral administration of short hairpin RNA constructs to target and reduce progerin has also demonstrated efficacy in HGPS fibroblasts. Therefore, pharmacological targeting to correct cellular phenotypes associated with HGPS could be utilized in the future for therapeutic intervention.

Introduction

Hutchinson-Gilford progeria syndrome (HGPS) is a very rare genetic autosomal dominant condition characterized by the dramatic, rapid appearance of aging beginning in childhood. Children with HGPS have characteristic craniofacial abnormalities and exhibit retarded growth, osteoporosis, osteolytic bone lesions, alopecia, wrinkled/aged skin due to loss of subcutaneous adipose tissue and vascular disease, which generally leads to death during the teenage years. This condition results from

mutations in the *LMNA* gene that produces a protein called prelamin A, which provides structural and mechanical stability for the nuclear envelope.

Therapy for individuals with HGPS is currently symptomatic and supportive. To date, no specific pharmacological agents have been identified for this genetic disorder, which is estimated to affect approximately 1 child in 4 million (1). However, recent studies have identified new targets and potential therapeutic strategies. Table I presents the therapeutic candidates for HGPS, which are discussed in more detail below.

Targets and therapeutic advances

Approximately 80% of HGPS cases carry a single nucleotide substitution at position 1824, C > T (G608G), within exon 11 of LMNA (GGC > GGT), and abnormal splicing results in the output of a truncated protein product with 50 amino acids deleted near its carboxy terminus. This mutant prelamin A is commonly known as progerin (2). Farnesylation, one of the enzymatic posttranslational modifications involved in the generation of mature lamin A, is important for the targeting of prelamin A to the nuclear envelope (3). Although truncation of progerin prevents the production of mature lamin A, it is thought that progerin does not side-step farnesylation and is thus transferred to the nuclear envelope, where it adversely affects the integrity of the nuclear lamina, resulting in nuclear deformations, blebbing and loss of peripheral heterochromatin (4).

Several studies have hypothesized that blocking farnesylation of progerin via the use of farnesyltransferase inhibitors, developed to treat cancer, may prevent its targeting to the nuclear envelope and reduce nuclear blebbing. Application of the farnesyltransferase inhibitor FTI-277 has been shown to decrease the proportion of abnormal nuclei from 31% to 8% (5). Further investigation of other farnesyltransferase inhibitors, including the clinical candidate lonafarnib (Sch-66336, Sarasar™), FTI-2153 and L-744832, indicated that these agents restore normal nuclear architecture in progerin-transfected HeLa, HEK-293 and NIH/3T3 cells and successfully reduce nuclear blebbing in human early- and late-stage HGPS fibroblasts (6). The farnesyltransferase inhibitor lovastatin in combination with the histone deacetylase

Table I: Chemical structures of the Drug	Structure	Ref.
FTI-277	H ₂ N CH ₃	5
Lonafarnib	Br CI NH ₂	6
FTI-2153	H ₃ C CH ₃	6
L-744832	H ₂ N CH ₃ CCH ₃ CCH ₃	6
Lovastatin + Trichostatin A	H_3C CH_3	7
ABT-100	NC OH CN OH N CH ₃	8

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inhibitor trichostatin A also rescued heterochromatin organization and transcript distribution, effects that were associated with a dramatic reduction in progerin levels in a small sample of fibroblasts obtained from HGPS patients (7).

In vivo assessment of the therapeutic potential of farnesyltransferase inhibition was carried out in an animal model of mandibuloacral dysplasia, a progeroid disorder similar to HGPS, which is caused by deficiency of the zinc mellanoproteinase ZMPSTE24, which cleaves the prelamin A precursor and its farnesyl lipid anchor to generate lamin A. ABT-100 (hydrochloride) improved body weight, grip strength, bone integrity and survival (8).

Overall, it appears that preventing farnesylation of progerin may be an effective strategy to reduce HGPS pathogenicity (3, 9, 10).

A recent study utilized gene therapy to correct cellular phenotypes associated with HGPS. Short hairpin RNA constructs, developed to target the mutated progerin mRNA, were applied to HGPS fibroblasts via lentiviral transmission. This RNA interference technology reduced progerin expression by 26%, with an associated normalization of nuclear morphology, improvement in proliferative potential and a reduction in senescent cells (11).

Some studies also suggest that the symptoms of HGPS may be due to the production of a biologically inactive form of growth hormone, accompanied by an elevated basal metabolic rate. A small study that applied nonaggressive nutritional therapy with growth hormone treatment indicated that this combination improved growth velocity, increased growth factor levels and decreased the basal metabolic rate, although these effects were not maintained and did not prevent the progression of vascular disease (12).

The National Institutes of Health Clinical Center is conducting a clinical investigation into HGPS. This study will examine which body systems are affected in progeria and how each system is affected over time in order to develop new treatments (13).

References

- 1. Pollex, R.L., Hegele, R.A. *Hutchinson-Gilford progeria syndrome*. Clin Genet 2004, 66(5): 375-81.
- 2. Eriksson, M., Brown, W.T., Gordon, L.B. et al. Recurrent de novo point mutations in lamin A cause Hutchinson-Gilford progeria syndrome. Nature 2003, 423(6937): 293-8.

- 3. Toth, J.I., Yang, S.H., Qiao, X. et al. *Blocking protein farnes-yltransferase improves nuclear shape in fibroblasts from humans with progeroid syndromes*. Proc Natl Acad Sci USA 2005, 102(36): 12873-8.
- 4. Goldman, R.D., Shumaker, D.K., Erdos, M.R. et al. *Accumulation of mutant lamin A causes progressive changes in nuclear architecture in Hutchinson-Gilford progeria syndrome*. Proc Natl Acad Sci USA 2004, 101(24): 8963-8.
- 5. McClintock, D., Gordon, L.B., Djabali, K. *Hutchinson-Gilford* progeria mutant lamin A primarily targets human vascular cells as detected by an anti-lamin A G608G antibody. Proc Natl Acad Sci USA 2006, 103(7): 2154-9.
- 6. Capell, B.C., Erdos, M.R., Madigan, J.P. et al. *Inhibiting far-nesylation of progerin prevents the characteristic nuclear blebbing of Hutchinson-Gilford progeria syndrome*. Proc Natl Acad Sci USA 2005, 102(36): 12879-84.
- 7. Columbaro, M., Capanni, C., Mattioli, E. et al. Rescue of heterochromatin organization in Hutchinson-Gilford progeria by drug treatment. Cell Mol Life Sci 2005, 62(22): 2669-78.
- 8. Fong, L.G., Frost, D., Meta, M., Qiao, X., Yang, S.H., Coffinier, C., Young, S.G. *A protein farnesyltransferase inhibitor amelio-rates disease in a mouse model of progeria*. Science 2006, 311: 1621-3.
- 9. Yang, S.H., Bergo, M.O., Toth, J.I. et al. *Blocking protein far-nesyltransferase improves nuclear blebbing in mouse fibroblasts with a targeted Hutchinson-Gilford progeria syndrome mutation.*Proc Natl Acad Sci USA 2005, 102(29): 10291-6.
- 10. Glynn, M.W., Glover, T.W. Incomplete processing of mutant lamin A in Hutchinson-Gilford progeria leads to nuclear abnormalities, which are reversed by farnesyltransferase inhibition. Hum Mol Genet 2005, 14(20): 2959-69.
- 11. Huang, S., Chen, L., Libina, N., Janes, J., Martin, G.M., Campisi, J., Oshima, J. Correction of cellular phenotypes of Hutchinson-Gilford progeria cells by RNA interference. Hum Genet 2005, 118(3-4): 444-50.
- 12. Abdenur, J.E., Brown, W.T., Friedman, S., Smith, M., Lifshitz, F. Response to nutritional and growth hormone treatment in progeria. Metabolism 1997, 46(8): 851-6.
- 13. Clinical studies of progeria (NCT00094393). ClinicalTrials.gov Web Site 2005, September 6.

Online links

Subscribers to Drugs of the Future can access an online animation to illustrate the underlying genetic cause of HGPS.